



Quick Guide

Trichin-L

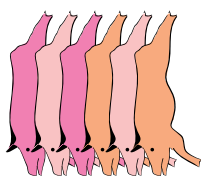
Trichinella Antigen Test Kit

357-2120

1. Collection and Preparation of the Sample

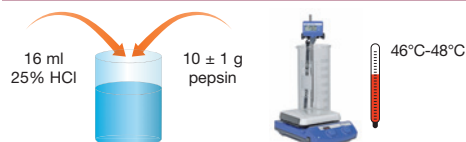
Tissue Sampling

x100
pillar
domestic swines



- **Domestic swine**
Pillar of the diaphragm 1 g to 1.15 g
Diaphragm, masseter or tongue 2 x 1 g
- **Breeding sows and boars**
Pillar of the diaphragm 2 x 1 g
Diaphragm, masseter or tongue 4 x 1 g

Preparation of the Digestion Buffer



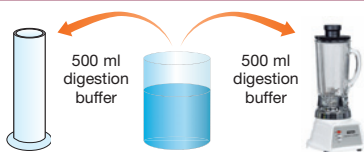
- Prepare 2 L of tap water pre-heated to 46-48°C in a 3 L beaker
- Add 16 ml of 25% HCl (0.2% final) and 10 ± 1 g of pepsin (0.5% final)
- Incubate at 46-48°C while stirring

Meat Chopping



- Add 100 - 115 g of pooled meat and 150 ml of digestion buffer (1:1.5 mass/vol.)
 - Mix 20 seconds at 18,000 rpm (speed 1)
- Note: only 10 seconds for prechopped samples used for ring trials*

Pepsin Digestion



- Keep 500 ml of digestion buffer in a graduated cylinder
- Pour 500 ml of digestion buffer into the blender bowl

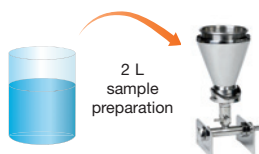


- Pour sample homogenate (~750 ml) into the 3 L beaker



- Carefully rinse the blender bowl with 500 ml of digestion buffer kept in the graduated cylinder. Pour the rinsing liquid into the 3 L beaker
 - Incubate 30 minutes at 44-46°C while stirring
- Switch ON the filtration pump few seconds before stopping the sample digestion**

Filtration



- Pour the sample preparation (2 L) into the filtration ramp through the filtration membrane

Note: rinse the 3 L beaker with at least 250 ml of warm water

Antigen Solubilization



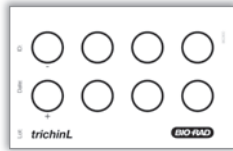
- Using forceps, remove the filtration membrane from the filtration ramp. Fold it in four then push it at the bottom of a 15 ml Falcon® tube with the pestle
- Crush the filtration membrane with the pestle (20 back and forth movements)
- Add 500 µl of sample diluent (R2). Thoroughly mix the *Trichinella* antigens with short back and forth movements (30 seconds)

Reading this information sheet is no substitute for reading the kit instructions.



2. Detection

Dispensing Samples & Controls



- **Samples & controls**

- 2 drops (50 µl) of negative control in well “-”
- 2 drops (50 µl) of positive control in well “+”
- 50 µl of sample #1 in well #1
- 50 µl of sample #2 in well #2, etc.

- **Conjugate**

- 1 drop (25 µl) of latex beads (R7) in each well
- Mix the conjugate and sample/control drops with a stick

Note: Carefully homogenize latex beads before use (minimum 10 tube inversions or light vortexing)

Rocking



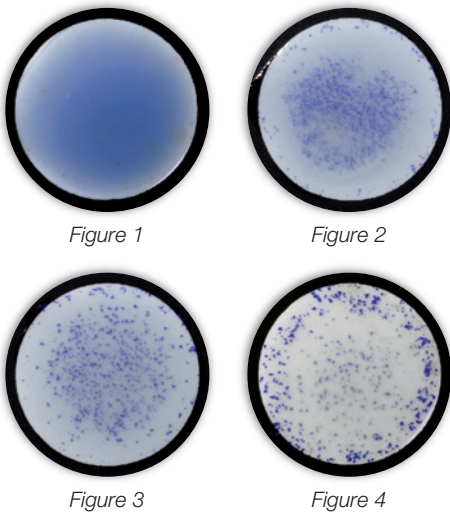
- Start rocking 10 minutes at maximum speed (~30 rpm)

Reading



- Stop rocking and immediately read the agglutinations

Interpretation of Results



- **Negative reaction:** the suspension remains blue and homogenous (see figure 1). Compare with the negative control
- **Positive reaction:** formation of bead aggregates (see figures 2, 3 and 4). The sample contains *Trichinella* antigens.

See *Trichin-L* kit insert for further details on the interpretation of results.

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